

0960-894X(94)E0077-R

ALKYL ETHER ANALOGS OF THE FK-506 RELATED, IMMUNOSUPPRESSIVE MACROLIDE L-683,590 (ASCOMYCIN)

Mark T. Goulet,** Derek W. Hodkey,* Mary Jo Staruch, Francis J. Dumont, John G. Cryan, William H. Parsons,* Matthew J. Wyvratt*

Abstract: C32-O-Ether derivatives of L-683,590 have been prepared by alkylation with various alkyl, alkenyl, and alkynyl 2,2,2-trichloroacetimidates. These analogs exhibit good immunosuppressive activity in vitro, as measured in a T cell proliferation assay. In the case of cinnamyl ethers, this activity correlates with the lipophilicity (π value) of the aromatic substituents.

The clinical importance of a potent and safe immunosuppressant is becoming increasingly evident as the present therapeutic agents, notably cyclosporin A (CsA)¹ and FK-506², find utility in the treatment of diseases beyond organ transplant rejection (e.g., psoriasis, rheumatoid arthritis, etc).³ It is thought that most autoimmune

diseases can be treated effectively with immunosuppressants, provided the safety profile of such agents would permit long term administration. Along with organ transplantation, FK-506 1 is currently under clinical evaluation for several autoimmune indications; however, it is not clear whether the neuro- and nephrotoxicity exhibited by this drug will allow for use in the treatment of chronic autoimmune diseases.⁴

CsA and FK-506 act to suppress the immune system by inhibiting the proliferation of antigen stimulated T cells. While they block the same calcium-dependent signal transduction pathway in T cells⁵, FK-506 is 100-fold more potent in vitro.⁶

			TOUR (IC50, RM)
1	FK-506	allyl	0.29
2	L-683,590	ethyl	0.69

The goal of our research is to discover semi-synthetic analogs of FK-506 or the structurally related L-683,590 (ascomycin)⁷ 2 that maintain the superior activity of this class of immunosuppressants while displaying fewer side-effects. As part of an ongoing chemical derivatization effort, a series of C32-O-alkyl ether analogs of L-683,590 were prepared with the object of modulating the lipophilicity of this compound.

Chemistry

Etherification of the hydroxyl groups in compounds such as L-683,590 requires a method that is non-basic and thermally mild to avoid complicating side-reactions.⁸ The acid catalyzed reaction of alkyl 2,2,2-trichloroacetimidates with alcohols provides the corresponding ethers at room temperature and under essentially neutral conditions.⁹ This procedure has been used to couple a variety of alkyl groups with alcohols, and has been employed extensively as a mild and reliable means of delivering alcohol protecting groups. We have found this method to be a convenient one for the preparation of ether derivatives of L-683,590 and related compounds.¹⁰ As shown in Scheme 1, treatment of 2 with allyl 2,2,2-trichloroacetimidate^{9b} (2 equiv.) and trifluoromethanesulfonic acid (TfOH, 0.2 equiv.) in cyclohexane/methylene chloride (2:1) provided a mixture of C32-O-allyl- and bis(C24, C32-O-allyl)-L-683,590, 3 and 4, in 78% yield.¹¹

Scheme 1

A modest degree of regioselectivity favoring alkylation of the C32 hydroxyl group is observed in these transformations; however, in many cases it was found desirable to protect the competing C24-OH as the t-butyldimethylsilyl ether prior to alkylation. This was accomplished by the two-step, bis-silylation/mono-desilylation protocol depicted in Scheme 2. Alkylation followed by deprotection of the C24-OTBS group by one of the hydrogen fluoride-mediated methods produced the C32-O-ether analogs.

Scheme 2

The reactivity of an alkyl 2,2,2-trichloroacetimidate is dependent on the ability of the alkyl group to support an incipient partial positive charge along the reaction coordinate. For instance, p-methoxybenzyl 2,2,2-trichloroacetimidate is a very reactive alkylating agent^{9d}, whereas we have found that reagents such as 6 and 7 do not react under the normal conditions. In the latter cases, prolonged exposure with L-683,590 and TfOH gave the

trichloroacetimidate-transfer product 8.¹² Thus, our alkylations are limited to the use of benzylic, allylic, or other non-primary, alkyl 2,2,2-trichloroacetimidate reagents.

In the case of certain allylic 2,2,2-trichloroacetimidates, there exists an issue of alkylation regiochemistry. When alcohol 5 was treated with various, substituted cinnamyl 2,2,2-trichloroacetimidate reagents 9 under the standard conditions, ether adducts arising from both α - and γ -alkylation, 10 and 11, were obtained (Scheme 3). The α -alkylation pathway was generally favored in the cinnamyl series (1-3:1), however, aromatic substituents were found to influence this ratio, with electron-withdrawing groups (X in 9 = 4-F, -Cl, -Br) resulting in a greater proportion of α -product. Cinnamyl derivatives such as 10 could be further modified by hydrogenation (H₂, Rh/C cat., EtOH) to provide the corresponding phenpropyl ethers.

Scheme 3

Results and Discussion

A series of C32-O-ether derivatives of L-683,590 were prepared to examine the effect of this modification on immunosuppressant activity (Table 1). Simple alkyl, alkenyl, and alkynyl ether derivatives (12-14) exhibit an approximate ten-fold loss in potency in a T cell proliferation assay¹³ compared to the parent alcohol L-683,590. The benzyl ether derivative 15 was found to be a marginally better immunosuppressant *in vitro*, with this activity being sensitive to α-branching (16). Two heteroaromatic analogs (17, 18) were found to be less active than their benzyl counterpart. Several 3'-substituted allyl ethers were also examined (19-22). While general alkyl and alkenyl substitution at this site did not improve activity relative to allyl ether 13, the cinnamyl ether analog 22 exhibited a demonstrable (4-fold) gain in potency. Several cinnamyl-like ethers containing modified aromatic tethers were prepared to investigate the role of the *trans* propenyl linking chain (23-25). These were found to possess slightly inferior immunosuppressive activity compared to 22, suggesting that among this series the propenyl tether is optimum.

An investigation of the C32-O-cinnamyl ether derivatives of L-683,590 was conducted in an attempt to increase the potency of this class. It was found that varying the substituents (X) on the aromatic ring resulted in an 18-fold range of *in vitro* activities, Table 2. Halogen substitution (26-30) was found to have an adverse

Table 1: In vitro immunosuppressive activity of C32-O-ether analogs of L-683,590

Compound	Ether group (R)	T Cell ¹³ IC ₅₀ (nM)
2	Н	0.7
12	Me Me	12.5
13	~	7.4
14	Мо—	6.1
15		3.0
16	—————————————————————————————————————	11.3
17		13.9
18		7.1
19	Me Me Me	14.2
20	Mo Mo	6.9
21		5.5
22	Q~	1.9
23		5.5
24	<u></u>	2.7
25		3.5

effect on potency relative to the parent cinnamyl ether 22, whereas, incorporation of a hydroxyl substituent at the 3- or 4- position (33, 34) served to increase immunosuppressive activity to a level approximating that of the parent natural product L-683,590, 2.

Analysis of the data from this study reveals a striking correlation between the *in vitro* potency of these analogs and the π -value ¹⁴ associated with the aromatic substituent. As shown in Figure 1, analogs containing substituents with low π -values are much more active as immunosuppressants than those with the highest π .

Table 2: In vitro immunosuppressive activity of C32-O-cinnamyl ether analogs of L-683,590

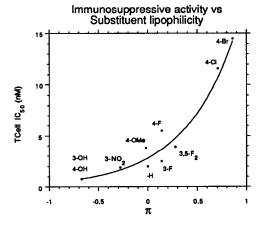
Compound	X-	π-value ¹⁴	T Cell ¹³ IC ₅₀ (nM)	FKBP12 ¹⁶ EC ₅₀ (nM)
26	4-Br	0.86	14.5	23.0
27	4-Cl	0.71	11.6	15.8
28	3,5-F ₂	0.28	3.9	
29	4-F	0.14	5.5	
30	3-F	0.14	2.5	
22	Н	0.0	1.9	29.7
31	4-OMe	-0.02	3.8	
32	3-NO ₂	-0.28	1.9	34.3
33	3-OH	-0.67	0.8	
34	4-OH	-0.67	0.8	20.5
L-683,590 2	••		0.7	1.6

Further, a correlation exists amongst the entire group, where a non-linear relationship is observed between substituent π -value and potency. A connection between bioactivity, both *in vitro* and *in vivo*, and compound lipophilicity has been observed within many drug classes.¹⁵ While other factors (e.g., electronic, steric) may also

be operating in this system, the observed correlation of immunosuppressive activity with substituent π -value suggests that the lipophilicity of this region exerts a significant influence on *in vitro* potency.

It is of interest that the *in vitro* immunosuppressive activity of these compounds does not directly correlate with their ability to bind the major cytosolic receptor, FKBP12, as measured in a competitive binding assay, Table 2.¹⁶ For example, analogs 26 and 34 bind to FKBP12 with equal affinities (ca. 20 nM), while their immunosuppressive activities differ 18-fold. Further, a 13-fold difference exists in the binding EC50 values of 34 and 2, yet they possess equivalent immunosuppressive activity. The enhanced

Figure 1:



immunosuppressive activity of L-683,590 analogs such as 34 is perhaps due to an alteration in cellular penetration, intracellular localization, or even the establishment of a new and favorable interaction with the target protein for this system, calcineurin. 17,10

In summary, a series of structurally diverse C32-O-alkyl ether analogs of L-683,590 were prepared and evaluated as immunosuppressants in an in vitro T cell proliferation assay. This study resulted in the identification of a class of C32-O-cinnamyl ether analogs with in vitro potency essentially equivalent to that of the parent natural product. The in vivo evaluation of some of these analogs for immunosuppressive efficacy and toxicity will be the subject of a future report.

The authors thank Dr. G. Salituro and Mr. F.P. Gailliot for providing the L-683,590 used Acknowledgment: in this study, and Dr. L.Colwell and Ms A. Bernick for mass spectrometry support.

References and Notes

- (a) Borel, J.F.; Feurer, C.; Gubler, H.U.; Stahelin, H. Agents Actions 1976, 468. (b) Wenger, R.M. 1. Angew. Chem. Int. Ed. Eng. 1985, 24, 77.
- Angew. Cnem. Int. Ea. Eng. 1985, 24, 1/.

 (a) Kino, T.; Hatanaka, H.; Hashimoto, M.; Nishiyama, M.; Goto, T.; Okuhara, M.; Kohsaka, M.; Aoki, H.; Imanaka, H. J. Antibiot. 1987, XL, 9, 1249 and 1256. (b) Tanaka, H.; Kuroda, A.; Marusawa, H.; Hatanaka, H.; Kino, T.; Hashimoto, M. J. Am. Chem. Soc. 1987, 109, 5031.

 Parsons, W.H.; Sigal, N.H.; Wyvratt, M.J. "Immunomodulating Drugs" Annals of the New York

 Acadamy of Sciences 1993, 685, 22-36. 2.
- 3.
- (a) Thomson, A.W.; Carroll, P.B.; McCauley, J.; Woo, J.; Abu-Elmagd, K.; Starzl, T.E.; Van Thiel, D.H. Springer Semin. Immunopathol. 1993, 14, 323.(b) Frank, B.; Perdrizet, G.A.; White, H.M.; Marsh, J.W.; Lemann, W.; Woodle, E.S. Transplant. Proc. 1993, 25, 1887. 4.
- Schreiber, S.L., Crabtree, G.R. Immunology Today, 1992, 13, 136.
- Sawada, S.; Suzuki, G.; Kawase, Y.; Takaku, F. J. Immunol. 1987, 139, 1797.
- Byrne, K.M.; Shafiee, A.; Nielsen, J.B.; Arison, B.; Monaghan, R.L.; Kaplan, L. In Microbial Metabolites; Nash, C., Ed.; Developments in Industrial Microbiology Series, Vol. 32; Wm. C. Brown: Dubuque, 1992, pp. 29-45.
- (a) Ok, H.; Arison, B.H.; Ball, R.G.; Beattie, T.R.; Fisher, M.H.; Wyvratt, M.J. Tetrahedron Lett. 8. 1990, 31, 6477. (b) Askin, D.; Reamer, R.A.; Jones, T.K.; Volante, R.P.; Shinkai, I. Tetrahedron Lett. 1989, 30, 671.
- (a) Iversen, T.; Bundle, D.R. J. Chem. Soc., Chem. Commun. 1981, 1240. (b) Wessel, H.-P.; 9. Iversen, T.; Bundle, D.R. J. Chem. Soc., Perkin Trans. 1 1985, 2247. (c) Armstrong, A.; Brackenridge, I.; Jackson, R.F.W.; Kirk, J.M. Tetrahedron, Lett. 1988, 29, 2483. (d) Nakajima, N.;
- Horita, K.; Abe, R.; Yonemitsu, O. *Tetrahedron, Lett.* 1988, 29, 4139. Goulet, M.T.; Hodkey, D.W.; Staruch, M.J.; Dumont, F.J.; Lin, S.; Hung, S.H.Y.; Siekierka, J.J.; Wyvratt, M.J. *BioMed. Chem. Lett.* 1994, 4, 0000, accompanying paper. 10.
- Satisfactory ¹H NMR and mass spectral data were obtained on all reaction products. This phenomenon has been reported previously and termed "reimidation": Danklmaier, J.; Honig, H. Liebigs Ann. Chem. 1989, 665-669. It is noteworthy that reagents such as 6 and 7 could serve a useful purpose for the acid-catalyzed formation of 2,2,2-trichloroacetimidates from alcohols, complementing the standard base promoted synthesis. An attempt to use 8 itself as an alkylating agent (allyl alcohol, TfOH (cat.), C₆H₁₂/CH₂Cl₂) gave no reaction.
- Assay was conducted using murine splenic T cells activated with PMA and ionomycin. In all cases the 13. inhibition observed was reversed by the addition of exogenous IL-2, see:
- Dumont, F.J.; Staruch, M.J.; Koprak, S.L.; Melino, M.R.; Sigal, N.H. J. Immunol. 1990, 144, 251. Fujita, T.; Iwasa, J.; Hansch, C. J. Am. Chem. Soc. 1964, 86, 5175. π is defined: $\pi = \log P_X - \log P_H$; 14. where: $P_{\rm H}$ is the octanol-water partition coefficient of the parent compound, and $P_{\rm X}$ is the partition coefficient of the derivative.
- 15. (a) Hansch, C.; Dunn, W.J. J. Pharm. Sci. 1972, 61, 1, and ref. cited therein. (b) Hansch, C.; Clayton, J.M. J. Pharm. Sci. 1973, 62, 1, and ref. cited therein.
- Siekierka, J.J.; Hung, S.H.; Poe, M.; Lin, C.S.; Sigal, N.H. Nature 1989, 341, 755. 16.
- 17. Liu, J.; Farmer, J.D.; Lane, W.S.; Friedman, J.; Weissman, I.; Schreiber, S.L. Cell 1991, 66, 807.